

Circulating Markers in Squamous Cell Carcinoma of the Head and Neck: A Review

Christopher H. Rassekh, Jonas T. Johnson and David E. Eibling

Biological markers of disease enhance the ability to diagnose, treat and evaluate results of therapy and are especially intriguing for their potential use in the management of malignant tumours. The serum levels of various biochemical substances have been shown to be abnormal for many cancers and are utilised in the management of affected patients. Several markers have been thoroughly investigated for potential clinical utility in head and neck carcinoma. Although no single marker has been found to be adequately sensitive and specific, combinations of markers may improve the utility for some aspects of patient management. This review highlights the literature to date in the realm of circulating markers for head and neck carcinoma. A discussion of the potential usefulness and limitations of such markers follows.

Oral Oncol, Eur J Cancer, Vol. 30B, No. 1, pp. 23-28, 1994.

INTRODUCTION

AN ESTIMATED 42400 new cases of cancer of the upper aerodigestive tract will be reported in the U.S.A. in 1993. The majority of these cases are squamous cell carcinoma (SCC) of the larynx, with oral cavity carcinoma a close second. Despite improvements in therapeutic and reconstructive modalities, head and neck carcinoma represents an important cause of cancer morbidity and mortality. The 5-year survival rates for malignancy of the oral cavity and pharynx remains lower than 55% and the head and neck is the only anatomical region in which 5-year survival rates have not improved significantly in the past two decades. It is estimated that over 2000 people will die from oral cancer alone in 1993 [1].

Detection of these malignancies is often delayed until advanced stage disease is present, since early lesions often do not produce symptoms. This is true for recurrent or second primary tumours as well as primary tumours. Monitoring response to therapy for carcinoma in this region is limited to techniques such as radiographic investigation and close clinical observation. The development of a reliable circulating tumour marker could assist in the evaluation of patients, by potentially expediating the detection of occult malignancy and could be used to monitor therapy. In addition, such a marker

might be used to develop new forms of treatment and might predict the biological behaviour of a tumour.

The ideal tumour marker would be a sensitive indicator of disease such that a high percentage of patients would have serum levels above the established normal. It would also be specific in that normal controls would not have levels above the accepted standard. Serum concentrations would correlate with tumour burden and clinical outcome and would return to normal with successful therapy and reappear prior to clinical recurrence. The ideal marker would also be stable enough to be assayed in low concentrations at a cost which is not prohibitive [2].

Unfortunately, no such "ideal" marker has been identified, however, there are several markers which have been found to be worthy of study in head and neck carcinoma. These include oncofetal proteins and other proteins, enzymes, hormones, metabolic byproducts such as erythrocyte polyamines, immune parameters, lipids, oncolipids and gangliosides, viral markers, tumour-associated antigens, prostaglandins and prostacyclins, and base elements (Table 1).

The following is a review of the most important markers that have been investigated in head and neck carcinoma.

CARCINOEMBRYONIC ANTIGEN

Carcinoembryonic antigen (CEA) was first described by Gold and Freeman [3] as a tumour-specific antigen for colorectal cancer. The subsequent reports indicating that CEA is a marker for gastrointestinal malignancies [4] were followed by the finding that elevated levels of CEA were present in a multitude of malignancies and levels of CEA returned to normal after successful therapy [4, 5]. In addition, elevation of circulating CEA has been associated with various non-malignant conditions including inflammatory diseases of the bowel, pancreas and liver and benign lung disease, as well as uraemia and chronic cigarette smoking [6]. Silverman et al. studied CEA levels of the 276 patients with head and neck

Correspondence to J.T. Johnson.

C.H. Rassekh, Fellow in Advanced Head and Neck Oncologic Surgery at the Department of Otolaryngology at the University of Pittsburgh School of Medicine; J.T. Johnson is Professor, Departments of Otolaryngology and Radiation Oncology, Vice Chairman, Department of Otolaryngology, and Director, Division of Oncology and Immunology at the University of Pittsburgh School of Medicine, The Eye and Ear Institute, Suite 500, 203 Lothrop Street, Pittsburgh, Pennsylvania 15213; and D.E. Eibling is Chief, Section of Otolaryngology at the Veterans Administration Medical Center in Oakland, and Associate Professor at the Department of Otolaryngology at the University of Pittsburgh School of Medicine, Pittsburgh, Pennsylvania.

Received 24 Feb. 1993; provisionally accepted 26 Feb. 1993; revised manuscript received 30 Mar. 1993.

Table 1. Tumour markers investigated in head and neck carcinoma

Onsoferal protoins (2, 6, 7, 17, 40, 50, 52-54) Metabolic byproducts (2, 5, 7, 30, 31) CEAR Immune complexes (2, 5, 11, 25, 25, 54) Immune complexes (2, 5, 30, 31) Proteins Proteins (2, 5, 11, 25, 25, 54) Immune complexes (2, 25, 30, 31) Proteins (2, 2, 31, 32, 25, 54) Immune complexes (3, 31, 32, 32, 34) (3, 31, 32, 32, 34) Proteins (2, 2, 31, 32, 32, 34) Immune complexes (3, 31, 32, 32, 34) (3, 31, 32, 32, 34) Proteins (3, 31, 32, 32, 34) Immune complexes (3, 31, 32, 32, 34) (3, 31, 32, 32, 34) Proteins (3, 31, 32, 32, 34) Immune complexes (3, 31, 32, 32, 34) (3, 31, 32, 32, 34) Proteins (3, 31, 32, 32, 34) Immune complexes (3, 31, 32, 32, 34) (3, 32, 32, 32, 32) Obserpation (3, 31, 32, 32, 32, 32, 32, 32, 32, 32, 32, 32	Markers	References	Markers	References
2	Oncofetal proteins CEA		Metabolic byproducts Erythrocyte polyamines	[2, 29]
2, 8, 11, 25, 54, 54 Immune oppositions	Alpha fetoprotein	[2]	Immune parameters	13 25 30 311
Comparison Com	Proteins	[0 0 11 05 53 54]	Immunogiobuins Immine complexes	[2, 23, 30, 31]
2, 52-54	Ferritin 0	[2, 6, 11, 23, 32, 34] [2]	Cytokines and cytokine recentors (TNF, IL-2 recentor)	[33]
2, 12, 25 Immunostippressive acidic protein 147 Lipids	p-protein Tissue nolynentide antigen	[2, 52–54]	Immunosuppressive substance	[34, 35]
56 Lipids sinic acid (also protein-bound and free forms) 147 Lipid sinic acid (also protein-bound and free forms) 12, 14, 56 Oncolipids 20	8-2-microglobulin	[2, 12, 25]	Immunosuppressive acidic protein	[35]
47]	Ceruloplasmin	[26]	Linids	
2, 14, 56 Oncolipids Gangliosides coprotein 2, 14, 56 Gangliosides Gangliosides 2, 14 Gangliosides 2, 14 Cital markers 2, 14 Cital markers 2, 14 Cital markers 2, 14 Cital markers 3, 14 Cital markers 2, 14 Cital markers 4, 56 Italy Herpes simplex virus 14, 56 Tumour-associated antigen (same as CA 19-9) 17, 54 Tumour-associated antigen (same as CA 19-9) 18, 54 TaG-72) Captrointestinal carcinoma-ansociated antigen (same as CA 19-9) 19, 54 Cital Captrointestinal carcinoma antigen (same as CA 19-9) 10, 54 Cital Captrointestinal carcinoma antigen (same as CA 19-9) 11, 54 TaG-72) Cital Captrointestinal carcinoma antigen (same as CA 19-9) 18, 55 Cital Captrointestinal carcinoma antigen (same as CA 19-9) 19, 54 Cital Captrointestinal carcinoma antigen (same as CA 19-9) 10, 54 Cital Captrointestinal carcinoma antigen (same as CA 19-9) 11, 54 TaG-72) Cital Captrointestinal carcinoma antigen (same as CA 19-9) 12, 20-22 Captrointestinal carcinoma antigen (same as CA 19-9) 11, 54 Captrointestinal carcinoma antigen (same as CA 19-9) 12, 20-22 Captrointestinal carcinoma antigen (same as CA 19-9) 12, 21 Captrointestinal carcinoma antigen (same as CA 19-9) 13, 22-22 Captrointestinal carcinoma antigen (same as CA 19-9) 14, 56 Captrointestinal carcinoma antigen (same as CA 19-9) 15, 27 Captrointestinal carcinoma antigen (same as CA 19-9) 16, 54 Captrointestinal carcinoma antigen (same as CA 19-9) 17, 27 Captrointestinal carcinoma antigen (same as CA 19-9) 17, 27 Captrointestinal carcinoma antigen (same as CA 19-9) 18, 56 Captrointestinal carcinoma antigen (same as CA 19-9) 18, 56 Captrointestinal carcinoma antigen (same as CA 19-9) 19, 56 Captrointestinal carci	Transforming growth factor- α	[47]	Lipid sialic acid (also protein-bound and free forms)	[2, 25, 45–47, 49, 52, 55,
2, 14, 56 Oncolipids	Glycoproteins			[95]
2, 14	α-1-antitrypsin	[2, 14, 56]	Oncolipids	[2, 37]
2, 14 Viral markers tuning 2, 14 Viral markers tuning 2, 14 Viral markers tuning 2, 14 Epstein-Bart virus 14, 56 Tumour-associated artigens 14, 56 Tumour-associated artigens 17, 52, 60, 61 Gastrointestinal carcinoma antigen (same as CA 19-9) 17, 54 Tumour-associated artigens (CA-20, CA-125, CA 15-3, TAG-72) 17, 54 Tag-72 Castrointestinal carcinoma-associated artigens (CA-50, CA-125, CA 15-3, TAG-72) 17, 54 Tag-72 Castrointestinal carcinoma-associated artigens (CA-50, CA-125, CA 15-3, TAG-72) 17, 54 Tag-72 Castrointestinal carcinoma-associated artigens (CA-50, CA-125, CA 15-3, TAG-72) 17, 54 Tag-72 Castrointestinal carcinoma-associated artigens (CA-50, CA-125, CA	α-1-acid glycoprotein	[2, 14]	Gangliosides	[57]
2, 14 Epstein-Bart virus 12, 14 Epstein-Bart virus 14, 56 Herpes simplex virus 14, 56 Tumour-associated antigens 11, 52, 60, 61 Squamous cell cartinoma antigen 17, 52, 60, 61 Squamous cell cartinoma antigen 17, 54 Tumour-associated antigen (same as CA 19-9) 17, 54 Tumour-associated artigen (same as CA 19-3) Tumour-associated artigen (same as CA 19-3) Tumour-associated artigen (same as CA 15-3) Tumour-associat	α-2-HS glyocoprotein	[2, 14]	Viral markers	
14, 14 Herpes simplex virus 14, 56 Tumour-associated antigens 14, 56 Tumour-associated antigens 117, 52, 60, 61 Gaptamous cell carcinoma antigen 117, 52, 60, 61 Gaptamous cell carcinoma antigen 117, 54 Gaptamour-associated antigen (CA-50, CA-125, CA 15-3, TAG-72) 14 alkaline phosphatase 12, 20-22 Base element 12 Zinc 12 Z	Pre-albumin	[2, 14]	Epstein-Barr virus	[2]
17, 54 Tumour-associated antigens 17, 54 Tumour-associated antigens 17, 54 Gastrointestinal carcinoma autigen 17, 54 Gastrointestinal carcinoma autigen 17, 54 Gastrointestinal carcinoma autigen 17, 54 Tumour-associated antigen (same as CA 19-9) Tumour-associated trypsin inhibitor 18 19 Tumour-associated antigens (CA-50, CA-125, CA 15-3, TAG-72) TAG-72 Base element 18 19 TAG-72 Capper element 19 19 Copper 19 Capper element 19	Albumin	[2, 14]	Herpes simplex virus	[2]
17, 52, 60, 61 Squarous cell carcinoma antigen 17, 54 17, 54 17, 54 18, 54 18, 54 18, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19, 54 19,	Haptoglobin	[14, 50]	Timoir-accordated anticenc	
17, 54 Tumour-associated antigen (same as CA 19-9) 17, 54 Tumour-associated antigen (same as CA 19-9) 18, 54 Tumour-associated antigen (same as CA 19-9) 19, 54 Tumour-associated antigen (same as CA 19-9) 10, 54 Tumour-associated antigen (same as CA 19-9) 10, 54 Tumour-associated antigen (same as CA 19-9) 11, 52 Tumour-associated uppsin inhibitor 12, 20-22 Base element 13 TAG-72 14, 52 Tage 15 Tage 15 Tage 16 Tage 17, 57 Tage 18 Tage 18 Tage 19 Tage 19 Tage 10 Tage 11 Tage 12 Tage 13 Tage 14 Tage 15 Tage 15 Tage 16 Tage 17, 27 Tage 18 Tage 19 Tage 10 Tage 10 Tage 11 Tage 12 Tage 13 Tage 14 Tage 15 Tage 16 Tage 17, 27 Tage 18 Tage 19 Tage 10 Tage 10 Tage 10 Tage 10 Tage 11 Tage 12 Tage 13 Tage 14 Tage 15 Tage 15 Tage 16 Tage 17, 27 Tage 17, 27 Tage 18 Tage 19 Tage 10 Tage 10 Tage 10 Tage 11 Tage 12 Tage 13 Tage 14 Tage 15 Tage 15 Tage 16 Tage 17, 27 Tage 18 Tage 19 Tage 10 Tage 10 Tage 11 Tage 12 Tage 13 Tage 14 Tage 15 Tage 15 Tage 16 Tage 17, 27 Tage 18 Tage 19 Tage 10 Tage 10 Tage 11 Tage 11 Tage 12 Tage 13 Tage 14 Tage 15 Tage 15 Tage 16 Tage 17, 27 Tage 17, 27 Tage 18 Tage 19 Tage 10 Tage 10 Tage 11 Tage 12 Tage 13 Tage 14 Tage 1	I AG-72	[00]	I uniour associated antigens	10 30 43 40 EE 401
11, 54 Tuastromtestinate carcinoma-associated annigen (same as CA 19-9) Tuastromtestinate carcinoma-associated annigen (same as CA 19-9) Tag-72 Tag-73 Tag-74	CA 19-9	[17, 52, 60, 61]	Squamous celi carcinoma anugen	[2, 38–45, 49, 33, 02]
101 1 mour-associated trypsin minotor 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102 102	CA-50	[17, 54]	Gastrointestinal carcinollia-associated antigen (same as CA 19-9)	[17, 00, 01]
e phosphatase 2, 20-22 Base element 2	CA-125	[61]	1 umour-associated trypsin inhibitor	[6c]
e phosphatase al alkaline phosphatase al alkaline phosphatase sin D Copper [2, 50] Multiple markers [2, 23] CEA, LSA, SCC-Ag and CA-125 CEA, LSA, SCC-Ag and CA 19-9 CEA, LSA, SCC-Ag and TPA [2, 24, 25] CEA, LSA, SCC-Ag and TPA [2] CEA, SCC-Ag and TPA CEA, SCC-Ag and TPA [2] CEA, SCC-Ag and PA [2] CEA, SCC-Ag and PA [2] CEA, SCC-Ag and PA CEA, SCC-Ag and PA [2] CEA, SCC-Ag and PA CEA, SCC-Ag CE	CA 15-3	[10]	Unier cancer-associated anugens (CA-50, CA-125, CA 15-5, TAG-72)	
e phosphatase [2, 20–22] Base element at alkaline phosphatase [2] Zinc sin D Copper Copper [43] Copper [54] Aultiple markers [25] CEA, LSA, SCC-Ag and CA-125 CEA, LSA, SCC-Ag and LDH CEA, LSA, SCC-Ag and LDH [27] CEA, LSA, SCC-Ag and TPA [28] CEA, LSA, SCC-Ag and TAA [29] CEA, LSA, SCC-Ag and CA-125 CEA, LSA, SCC-Ag and CA-125 CEA, SCC-Ag and TPA [28] CEA, SCC-Ag and TPA [29] CEA, SCC-Ag and TPA [20] LSA and SCC-Ag and TPA [20] LSA and SCC-Ag and PHI PSA and SCC-Ag LSA, ferritin, P2-microglobulin, IgE and PHI PSA and SCC-Ag PSA and SCC-Ag Inin LSA, cert-acid glycoprotein [2] LSA, α-1-antitrypsin, haptoglobin and ceruloplasmin	Enzymes		(2, 231)	
alkaline phosphatase [2] Zinc n D Copper Copper lehydrogenase [2, 50] Multiple markers se [2, 23] CEA, LSA, SCC-Ag and CA-125 te elastase CEA, LSA, SCC-Ag and LDH hexose isomerase (2, 24, 25] CEA, LSA, SCC-Ag and LDH ne deaminase (2) CEA, LSA, SCC-Ag and LDH deoxyribonuclease [2] CEA, SCC-Ag and TPA deoxyribonuclease [2] CEA, SCC-Ag ferritin, TPA and CA-50 li7, 27] LSA and SCC-Ag li7, 27] LSA and SCC-Ag li7, 27] LSA and SCC-Ag lin PSA and SCC-Ag in (26) lin (27) lin (28) lin<	Alkaline phosphatase	[2, 20-22]	Base element	
n D [63] Copper 1ehydrogenase [2, 23] Multiple markers se [2, 23] CEA, LSA, SCC-Ag and CA-125 te elastase CEA, LSA, SCC-Ag and LDH hexose isomerase CEA, LSA, SCC-Ag and LDH ce deaminase CEA, LSA, SCC-Ag and TPA specific enolase CEA, SCC-Ag and TPA deoxyribonuclease [2] CEA, SCC-Ag and TPA deoxyribonuclease [2] LSA and SCC-Ag ine kinase LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and scc-Ag LSA, call and α-1-acid glycoprotein [2] LSA, α-1-antitrypsin, haptoglobin and ceruloplasmin [2] LSA, α-1-antitrypsin, haptoglobin and ceruloplasmin	Placental alkaline phosphatase	[2]	Zinc	[62]
1ehydrogenase [2, 59] Multiple markers se [2, 23] CEA, LSA, SCC-Ag and CA-125 te elastase [28] CEA, LSA, SCC-Ag and LDH hexxose isomerase [2] CEA, LSA, SCC-Ag and LDH ne deaminase [2] CEA, SCC-Ag and TPA specific enolase [2] CEA, SCC-Ag and TPA deoxyribonuclease [2] LSA and SCC-Ag ine kinase [26] LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag CEA and α-1-acid glycoprotein [2] CEA and α-1-acid glycoprotein [2] LSA, α-1-antitrypsin, haptoglobin and ceruloplasmin	Cathepsin D	[63]	Copper	[62]
se [2, 25] CEA, LSA, SCC-Ag and CA-125 te elastase [28] CEA, LSA, SCC-Ag and LDH hexose isomerase [27] CEA, LSA, SCC-Ag, ferritin, TPA and CA 19-9 ne deaminase [2] CEA, SCC-Ag and TPA specific enolase [2] CEA, SCC-Ag and TPA deoxyribonuclease [2] LSA and SCC-Ag ine kinase LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag CEA and α-1-acid glycoprotein [2] CEA and α-1-acid glycoprotein [2] LSA, α-1-antitrypsin, haptoglobin and ceruloplasmin	Lactate dehydrogenase	[2, 50]	Multiple markers	
te elastase [2, 24, 25] CEA, LSA, SCC-Ag and LDH hexose isomerase [2, 24, 25] CEA, LSA, SCC-Ag, ferritin, TPA and CA 19-9 ce deaminase [2] CEA, SCC-Ag and TPA ce deaminase [2] CEA, SCC-Ag and TPA ce deoxyribonuclease [2] CEA, SCC-Ag and CA-50 LSA and SCC-Ag in [2] LSA, ferritin, TPA and CA-50 LSA and SCC-Ag in [26] LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag in CEA and α-1-acid glycoprotein [27] CEA and α-1-acid glycoprotein [28] LSA, α-1-antitrypsin, haptoglobin and ceruloplasmin	Aliesterase	[2, 23]	CEA, LSA, SCC-Ag and CA-125	[49]
hexose isomerase[2, 24, 25]CEA, LSA, SCC-Ag, ferritin, TPA and CA 19-9ne deaminase[2]CEA, SCC-Ag and TPAspecific enolase[2]CEA, SCC-AG, ferritin, TPA and CA-50deoxyribonuclease[17, 27]LSA and SCC-Agine kinaseLSA, ferritin, β-2-microglobulin, IgE and PHIpSA and SCC-AgPSA and SCC-Agin[2]CEA and α-1-acid glycoproteinandins and prostacyclins[2]LSA, α-1-antitrypsin, haptoglobin and ceruloplasmin	Leukocyte elastase	[28]	CEA, LSA, SCC-Ag and LDH	[50]
specific enolase [2] Sec A, SCC-Ag and TPA [2] CEA, SCC-AG, ferritin, TPA and CA-50 Geoxyribonuclease [17, 27] LSA and SCC-Ag Inc kinase [26] LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag Inc Kinase [27] SSA and SCC-Ag Inc Kinase [28] Inc Kinase In	Phosphohexose isomerase	[2, 24, 25]	CEA, LSA, SCC-Ag, ferritin, TPA and CA 19-9	[52]
specific enolase [2] CEA, SCC-AG, ferritin, TPA and CA-50 deoxyribonuclease [17, 27] LSA and SCC-Ag ine kinase [26] LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag Incompare the strict of the strict	Adenosine deaminase	[2]	CEA, SCC-Ag and TPA	[53]
deoxyribonuclease [26] L.S.A. ferritin, β-2-microglobulin, IgE and PHI [26] L.S.A. ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag CEA and α-1-acid glycoprotein [2] CEA and α-1-acid glycoprotein [2] L.S.A. α-1-antitrypsin, haptoglobin and ceruloplasmin	Neuron-specific enolase	[2]	CEA, SCC-AG, ferritin, TPA and CA-50	[54]
LSA, ferritin, β-2-microglobulin, IgE and PHI PSA and SCC-Ag in [2] CEA and α-1-acid glycoprotein LSA, α-1-antitrypsin, haptoglobin and ceruloplasmin	Alkaline deoxyribonuclease	[17, 27]	LSA and SCC-Ag	[2, 55]
PSA and SCC-Ag in [2] CEA and α -1-acid glycoprotein and prostacyclins [2] LSA, α -1-antitrypsin, haptoglobin and ceruloplasmin	Thymidine kinase	[07]	LSA, ferritin, \beta-2-microglobulin, IgE and PHI	[2, 25]
(2) CEA and α -1-acid glycoprotein dins and prostacyclins [2] LSA, α -1-antitrypsin, haptoglobin and ceruloplasmin	Hormones		PSA and SCC-Ag	[51]
[2] LSA, a-1-antitrypsin, haptoglobin and ceruloplasmin	Calcitonin	[2]	CEA and α-1-acid glycoprotein	[2]
	Prostaglandins and prostacyclins	[2]	LSA, α -1-antitrypsin, haptoglobin and ceruloplasmin	[26]

carcinoma of whom 154 were smokers and 122 were nonsmokers. Both the incidence and magnitude of CEA elevations correlated with the clinical stage of disease; however, if patients with advanced disease were excluded similar levels of the antigen were found in tumour-bearing patients, tumourfree previously treated patients, and healthy controls. If the definition of abnormal CEA levels is 5 ng/ml, then 36% of tumour-bearing patients had abnormal levels whereas only 5% of the non-smoking control patient population had an abnormal level. If the definition of abnormal is a CEA greater than 7 ng/ml, 17% of the tumour-bearing patients and 5% of smoking control patients had abnormal levels. Schneider et al. [7] found that 47% of 85 patients with head and neck carcinoma had levels greater than 5 ng/ml but did not find a correlation with site or stage of disease and the level of the CEA. Currently, CEA is not felt to be a good marker for head and neck carcinoma.

PROTEINS AND GLYCOPROTEINS

Ferritin is a storage and major iron binding protein that is normally found in nanogram quantities in human serum. Elevated serum ferritin has been demonstrated in patients with lymphoma, leukaemia, multiple myeloma, breast cancer, lung cancer, colorectal cancer, liver cancer, malignant melanoma, neuroblastoma, tumours of the testes and ovaries and non-malignant conditions [8-10]. The potential for ferritin as a serum marker in head and neck cancer was investigated by Maxim and Veltri in 1986 [8]. These investigators found that there was a significant difference in serum levels between nonsmoking control patients and smoking control patients when compared to patients with head and neck cancer. The prognostic significance of persistently elevated ferritin levels during treatment was postulated. More recent studies suggest that early laryngeal cancers are not readily diagnosed due to a lack of specificity [11]. Thus, ferritin is not currently a useful marker for head and neck cancer.

Other proteins such as β -2-microglobulin and β -protein have been found to be elevated in serum of patients with head and neck carcinoma, however, the usefulness of these substances is unknown [2, 12]. Glycoproteins have been studied in various types of malignancies including those of the liver [13, 14]. Szymendera [15] showed that a variety of malignant tumours produced elevated levels of glycoproteins CA 19-9, CA-50 and CA-125. Wolf et al. [16] studied six glycoproteins: haptoglobins, α -1-antitrypsin, α -1-acid glycoprotein, α -2-HS glycoprotein, pre-albumin and albumin. They demonstrated a correlation between levels of α -1-antitrypsin and α -1-acid glycoprotein and tumour burden. Haptoglobin was elevated significantly in a group of tumour-bearing patients but no correlation between levels and tumour burden were demonstrated. The levels of α -2-HS glycoprotein decreased with increased tumour burden. More recent studies of glycoproteins have suggested that they are not adequate alone in head and neck tumour evaluation [17].

Fazekas-May [18] studied 14 patients with SCC of the head and neck and found that urinary transforming growth factor alpha levels did correlate, however, with the course of disease in 43% of the patients studied. This marker appears to warrant further study in head and neck carcinoma. However, at this time, measurement of this transforming growth factor (TGF- α) in head and neck cancer is not clinically useful.

SERUM ENZYMES

The level of various enzymes in serum has been correlated with different types of cancer. The best example of this is alkaline phosphatase which has been found to be present in elevated levels in hepatic carcinoma [19]. A study has been performed correlating response to chemotherapy for carcinoma of the head and neck with the pretreatment levels of alkaline phosphatase, [20] however, others presented data which did not agree with this finding [21, 22]. Aliesterase is an enzyme involved in fat metabolism and in contrast to enzymes required for glycolysis, aliesterase levels have been shown to decrease in patients with malignancy. One study in head and neck tumours revealed that serum levels of aliesterase varied inversely with tumour stage and that levels rose during radiotherapy but did not reach control levels [23]. Phosphohexose isomerase (PHI) is an enzyme that transforms glucose-6-phosphate and elevated levels have been found in patients with advanced tumours including head and neck cancers, [24] however, the marker is not sensitive enough to be useful in carcinoma of the head and neck [25]. Thymidine kinase was found to be elevated in the sera of oral carcinoma patients, but with a sensitivity of only 47% [26]. Alkaline deoxyribonuclease levels appear to correlate with successful treatment of head and neck cancer [27]. Another enzyme that was studied by Harbans Lal was serum adenosine deaminase which correlated with tumour burden and response to tumour therapy, however, these changes were not sufficiently specific to warrant their use as a diagnostic tumour marker [19]. Most recently, leukocyte elastase has been found to be elevated in a variety of pharyngeal cancers [28]. The usefulness of various enzyme markers in head and neck cancer requires further study. There is no apparent clinical utility of these markers currently for head and neck cancer.

ERYTHROCYTE POLYAMINES

Erythrocyte polyamines were studied in patients with head and neck cancer in 29 previously untreated patients of whom 31% had increased levels which correlated with tumour burden. The levels also decreased after surgery or irradiation [29]. These data suggest that this marker is also not sensitive enough for diagnostic purposes.

IMMUNE PARAMETERS

Abnormalities of the immune system are well documented in patients with head and neck carcinoma. Immunoglobulins, particularly IgE, and circulating immune complexes have been investigated, however, these are also not sensitive or specific enough to be valuable tumour markers [25, 30-32]. Recently, investigations of interleukin-2 (IL-2) receptor, tumour necrosis factor (TNF), immunosuppressive substance (IS) and immunosuppressive acidic protein (IAP) have begun which will probably change our understanding of these potential markers [33-35]. Hsu et al. found that soluble IL-2 receptor levels were elevated in patients with nasopharyngeal carcinoma and correlated with clinical staging, whereas levels of TNF were elevated, but did not correlate with tumour stage. IL-2 receptor is a blocking factor causing decreased mitogenic response, but its use as a marker requires further study [33].

Yamanaka et al. used a single radial immunodiffusion in 108 patients with head and neck cancer and found serum immuno

suppressive substance (IS) positive in 46%. They also found that elevation correlated with extent of disease and predicted recurrence [34]. Kubota et al. found 58% of patients with oral cancer had positive IS levels and found IS to be a useful parameter for monitoring disease stage and therapy [35]. These newer makers still lack adequate sensitivity to be useful as diagnostic aids.

ONCOLIPIDS

In 1986, Fossel et al. [36] described a new non-specific cancer marker involving the use of water suppressed proton nuclear magnetic resonance spectroscopy of plasma. Lipoprotein particles found in the plasma of cancer patients have been termed "oncolipids" and preliminary tests have suggested that this assay may provide a universal cancer screening test. One study [37] of 46 patients who had biopsy-proven head and neck carcinoma and 32 controls revealed significant differences between patients with disease and controls. However, the conclusion of this study was there was no clinical utility due to the tremendous overlap between disease and non-disease groups.

SQUAMOUS CELL CARCINOMA (SCC) ANTIGENS

A squamous cell-derived antigen TA-4 was initially isolated and purified from squamous cell carcinoma from the uterine cervix by Kato and Turagoe [38] in 1977 using a radioimmunoassay (RIA). Kato et al. demonstrated that serum levels of this antigen correlated with extent of disease as well as success of therapy [39]. Maruo et al. [40] demonstrated that elevated serum TA-4 levels fell very rapidly after complete tumour excision or following definitive radiation therapy. Studies of this TA-4 indicate that it is not a single substance but a series of at least 14 proteins with a common antigen and a molecular weight of approximately 48 000 daltons [40]. Initial studies demonstrated elevated SCC antigen in 13 out of 25 patients with head and neck carcinoma [41]. Johnson et al. [42] demonstrated elevated pretreatment SCC antigen levels in 45% of 60 patients with head and neck SCC. Eibling et al. found elevated pretreatment levels in 44% of 89 patients with head and neck carcinoma [43]. Although the mean SCC antigen level increased with increasing T or N stage of the disease, the percentage of patients with elevated levels did not vary significantly with increasing tumour burden. There was a strong correlation between serum antigen levels and clinical course in patients who developed recurrence. Until recently, this marker seemed to have the highest sensitivity of the markers studied in head and neck cancer. Although the sensitivity is still less than 50%, in those patients who present with elevated levels, post-therapy serum levels may provide clinically useful data. Routine clinical use of SCC-RIA is not recommended.

LIPIDS AND FORMS OF SIALIC ACID

Serum lipid-bound sialic acid (LSA) have been found to be useful in assessing disease progression in identifying patients resistant to therapy with breast and colorectal carcinoma although it is neither sensitive nor specific enough for cancer screening [25, 44, 45]. Fisher et al. [46] found that sialic acid levels in head and neck cancer patients were elevated in proportion to the extent of tumour and that 71% of patients

with recurrence of tumour had elevated levels. A previous study found the protein-bound form to be a better indicator, than LSA or free sialic acid of poor prognosis in head and neck malignancies [47]. The significance of these and other studies is discussed in the section on multiple markers. Sialic acid seems to be one of the more useful markers which should be investigated further, but still cannot be currently advocated as a diagnostic aid.

MULTIPLE MARKERS

The lack of a single tumour marker has led investigators to search for combinations of multiple markers. Gail et al. [48] found that the combination of CEA and total sialic acid were found to be useful in advanced lung cancer. Based on this model, Straka et al. [49] reviewed the use of a tumour marker panel and studied the utility of the combination of SCC antigen, CEA, LSA and CA-125. They confirmed that SCC antigen RIA was the most sensitive marker, however, various combinations of the markers did not significantly improve either specificity or sensitivity of the markers for diagnostic purposes. Dreyfus et al. [50] evaluated LSA, SCC antigen, CEA and lactate dehydrogenase (LDH) levels in squamous cell carcinoma of the head and neck in 52 patients. 42 of these patients had active measurable disease and 10 had no evidence of clinical disease. In patients with active disease, LSA, SCC antigen, CEA and LDH were elevated in 71, 33, 27 and 18%, respectively. None of the markers were elevated in the group with no evidence of disease. The incidence and magnitude of LSA and SCC antigen elevations correlated with the extent of disease and the authors concluded that LSA appeared to be the most promising and sensitive marker of SCC of the head and neck followed in decreasing order of sensitivity by SCC antigen, CEA and LDH. The difference between these two studies may be related to using a different assay method for LSA. Similarly, Ropka et al. found that the LSA test sensitivity was 63.4 vs. 27.6% for SCC-associated antigen with a specificity of 77.9% for LSA and 85% for SCCassociated antigen. When the combination of both tests were positive, sensitivity was 18.7% and specificity was 95%. If either was positive in parallel combinations, sensitivity was 72.4% and specificity was 68%. They concluded that further evaluation is required that applies different definitions of normal and determines longitudinal changes with disease status [51]. Vinzenz found elevated levels of β-2 microglobulin, IgE, PHI, LSA and ferritin in 50%, 55-62%, 62%, 60% and 50%, respectively, and concluded that LSA, β -2 microglobulin and ferritin merit further study for early detection [25]. Mevio et al. found LSA and ferritin to be more useful than CEA, TPA, SCC-Ag, CA19-9 in 50 patients with laryngeal cancer [52]. Palermo et al. found benefit from using the combination of SCC-Ag, CEA and TPA with an increased sensitivity of up to 71% [53].

Screm et al. studied CEA, ferritin, CA-50, TPA and SCC-Ag in 54 "ENT" cancers with a 72% combined sensitivity, and concluded that they were not useful since objective data is available even in early tumours [54]. Bhatavdekar also found PSA to be a more reliable marker than SCC antigen [55]. It would appear from all the data from these combined studies that sialic acid is the most sensitive marker for SCC, followed by SCC antigen and possibly ferritin. It would seem to follow that further investigations of these markers, and particularly the various forms of sialic acid is indicated. Combinations

including newer immune markers like IS should also be investigated further.

CONCLUSIONS

The identification of a circulating tumour marker has great potential for clinical application. Unfortunately, efforts to date to identify a single tumour marker or even a combination of markers for head and neck carcinoma are limited by an inadequate sensitivity and specificity. Therefore, serum markers are not widely used clinically and are primarily used in research. Despite this lack of a useful marker for diagnostic screening and recognition of occult malignancy, markers may be useful for monitoring therapy. The development of new multiple marker batteries may provide the closest approximation to a perfect marker that is feasible. Additional studies into the nature and function of oncogenes may provide clues to the variability of these tumours and the resultant heterogenieity of their marker expression.

- 1. Cancer Statistics. CA 1993, 43, 26.
- Eibling DE, Wagner RL, Johnson JT. Tumor markers of head and neck carcinoma. *Immunol Series* 1990, 53, 357–383.
- Gold P, Freedman SO. Demonstration of tumor-specific antigens in human colonic carcinomata by immunological tolerance and absorption techniques. J Exp Med 1964, 121, 439–471.
- 4. Staab HJ, Anderer FA, Stumpf E, Fischer R. Slope analysis of the postoperative CEA time course and its possible application as an aid in diagnosis of disease progression in gastrointestinal cancer. Am J Surg 1978, 136, 322–327.
- Reynoso G, Chu TM, Holyoke D, et al. Carcinoembryonic antigen in patients with different cancers. JAMA 1972, 220, 361-365.
- Silverman NA, Alexander JC, Chretien PB. CEA levels in head and neck cancer. Cancer 1976, 37, 2204–2211.
- Schneider M, Dermard F, Chauvel P, et al. Carcinoembryonic antigen determinations in head and neck cancer. In Krebs B, Lalanne C, Schneider M, eds. Clinical Applications of Carcinoembryonic Antigen Assay: Proceedings of a Symposium. Amsterdam, Excerpta Medica, 1978, 384–387.
- 8. Maxim PE, Veltri RW. Serum ferritin as a tumor marker in patients with squamous cell carcinoma of the head and neck. *Cancer* 1986, 57, 305–311.
- 9. Giler S, et al. The significance of ferritin in malignant disease. Biomedicine 1978, 28, 203-206.
- Luger TA, Linkesch W, Knobler R, Kokoschka EM. Serial determination of serum ferritin levels in patients with malignant melanoma. Oncology 1983, 40, 263–267.
- Tarchalska-Krynska B, Sawicka B, Zawisza E, Andrezejewski A. Ferritin (tumor marker) in patients with laryngeal cancer and precancerous conditions. Otolaryngologia Polska 1991, 45, 241–245.
- Manzar W, Raghavan MR, Aroor AR, Keshava Murthy KR. Evaluation of serum beta-2-microglobulin in oral cancer. Autr Dent J 1992, 37, 39-42.
- Harris CC, Primack A, Cohen MH. Elevated alpha₁-antitrypsin scrum levels in lung cancer patients. Cancer 1974, 34, 280–281.
- 14. Chio LF, Oon CH, Cantab. Changes in serum alpha₁ antitrypsin, alpha₁ acid glycoprotein and beta₂ glycoprotein I in patients with malignant hepatocellular carcinoma. Cancer 1979, 43, 569-604.
- Szymendera JJ. Clinical usefulness of three monoclonal antibodydefined tumor markers: CA 19-9, CA 50, and CA 125. Tumour Biol 1986, 7, 333–342.
- Wolf GT, Chretien PB, Elias EG, et al. Serum glycoproteins in head and neck squamous carcinoma: correlations with tumor extent, clinical tumor stage, and T-cell levels during chemotherapy. Am J Surg 1979, 138, 489-500.
- Gustafsson H, Franzen L, Grankvist K, Anniko M, Henriksson R. Glycoprotein tumor markers in head and neck neoplasms—a consecutive study on CA-50, CA 19-9, and CEA. J Cancer Res Clin Oncol 1988, 114, 394–398.
- Fazekas-May, M, Suen JY, Yeh YC, Yeh HW, Milligan LB. Investigation of urinary transforming growth factor alpha levels

- as tumor markers in patients with advanced squamous cell carcinoma of the head and neck. *Head Neck* 1990, 12, 411-416.
- Harbans Lal HC, Munjal SK, Wig U, Saini AS. Serum enzymes in head and neck cancer III. J Laryngol Otol 1987, 101, 1062-1065.
- Katz AE, Hong WK, Bhutani R, Berman LD, Blanchard GJ, Koff RS. Prognostic indicators in chemotherapy for head and neck carcinoma: alkaline phosphatase levels. *Laryngoscope* 1980, 90, 924-929.
- 21. Coker DD, Morris D, Elias G, Didolkar MS, Zentai TA. Head and neck cancer: relationship of the prechemotherapy serum alkaline phosphatase levels to response rate of induction chemotherapy. *Arch Otolaryngol* 1982, 108, 28–29.
- Burres SA, Jacobs JR, Peppard SB, Al-Sarraf M. Significance of alkaline phosphatase and chemotherapy for head and neck carcinoma. Otolaryngol Head Neck Surg 1982, 90, 188–192.
- Harbans Lal HC, Madan HC, Kohli GS, Yadav SPS. Serum enzymes in head and neck cancer II. J Laryngol Otol 1987, 101, 819-822.
- 24. Goel H, Kohli GS, Lal H. Serum phosphohexose isomerase levels in patients with head and neck cancer. J Laryngol Otol 1986, 100, 581-585
- Vinzenz K, Schönthal E, Zekert F, Wunderer S. Diagnosis of head and neck carcinomas by means of immunological tumour markers. J Cranio-Max-Fac Surg 1987, 15, 270-277.
- Scully C. Thymidine kinase activity in oral squamous cell carcinoma. J Oral Pathol 1982, 11, 210-213.
- Economidou-Karaoglou A, Opsomer M, Lans M, Taper HS, Deckers C, Roberfroid MB. Predictive value of serum alkaline DNase activity variations in treatment of head and neck cancer. Acta Oncologica 1990, 29, 163–166.
- Zoller E. The role of leukocyte elastase in malignant tumors of the head and neck. Laryngoscope 1989, 99, 971–973.
- Shideler CE, Johns ME, Cantrell RW, Shipe JR, Wills MR, Savory J. Erythrocyte polyamine determinations in patients with head and neck cancer. *Arch Otolaryngol* 1981, 107, 752–754.
- Katz AE, Nysather JO, Harker LA. Major immunoglobulin ratios in carcinoma of the head and neck. Ann Otol 1978, 87, 412–415.
- Veltri RW, Rodman SM, Maxim PE, Baseler MW, Sprinkle PM. Immune complexes, serum proteins, cell-mediated immunity, and immune regulation in patients with squamous cell carcinoma of the head and neck. Cancer 1986, 57, 2295–2308.
- Scully C, Barkas T, Boyle T, McGregor IA. Circulating immune complexes detected by binding of radiolabelled protein A in patients with oral cancer and premalignant lesions. J Clin Lab Immunol 1982, 8, 113-115.
- Hsu MM, Ko JY, Chang YL. Elevated levels of soluble interleukin-2 receptor and tumor necrosis factor in nasopharyngeal carcinoma. Arch Otolaryngol Head Neck Surg 1991, 117, 1257-1259.
- 34. Yamanaka N, Harabudri Y, Himi T, Katura A. Immunosuppressive substance in the sera of head and neck cancer patients. *Cancer* 1988, **62**, 1293–1298.
- Kubota E, Kurokawa H, Katsuki T. Evaluation of the serum level of immunosuppressive substance in oral cancer patients. J Oral Maxillofac Surg 1991, 49, 121-126.
- Fossel ET, Carr JM, McDonagh J. Detection of malignant tumors: water suppressed protein nuclear magnetic resonance spectroscopy of plasma. N Engl J Med 1986, 315, 1369–1376.
- Scher RL, Ropka ME, Neal DA, et al. NMR spectroscopy evaluation of plasma "oncolipids" in head and neck cancer. Otolaryngol Head Neck Surg 1990, 102, 3440.
- Kato H, Torigoe T. Radioimmunoassay for tumor antigen of human cervical squamous cell carcinoma. Cancer 1977, 40, 1621–1628.
- Kato H, Tamai K, Morioka H, Nagai M, Nagaya T, Torigoe T. Tumor-antigen TA-4 in the detection of recurrence in cervical squamous cell carcinoma. *Cancer* 1984, 54, 1544–1546.
- Maruo T, Shibata K, Kimura A, Hoshina M, Mochizuki M. Tumor-associated antigen, TA-4, in the monitoring of the effects of therapy for squamous cell carcinoma of the uterine cervix: serial determinations and tissue localization. Cancer 1985, 56, 302-308.
- Fukunga M, Otsuka N, Sone T, et al. Clinical study on the measurement of squamous cell carcinoma (SCC) related antigen in SCC. Gan No Rinsho 1985, 31, 1855-1888.
- 42. Johnson J, Wagner R, Eibling D, et al. Radioimmunoassay for

- SCC antigen in the diagnosis of squamous cell carcinoma of the head and neck: a preliminary report. In Kataloff, deBruijott, Ebert W, et al. eds. SCC Antigen in the Management of Squamous Cell Carcinoma. Princeton, Excerpta Medica, 1987, 112–123.
- Eibling DE, Johnson JT, Wagner RL. SCC-RIA in the diagnosis of squamous cell carcinoma of the head and neck. *Laryngoscope* 1989, 99, 117-124.
- 44. Dnistrian AM, Schwartz MK, Katopodis N, Francchia AA, Stock CC. Serum lipid-bound sialic acid as a marker in breast cancer. *Cancer* 1982, 50, 1815–1819.
- Erbil KM, Jones JD, Klee GG. Use and limitations of serum total and lipid-bound sialic acid concentrations as markers for colorectal cancer. Cancer 1985, 55, 404

 409.
- Fischer F, Egg G. N-acetyl neuraminic acid (sialic acid) as a tumor marker in head and neck cancer. HNO 1990, 38, 361–363.
- Bhatavdekar JM, Vara HH, Patel DD. Serum sialic acid forms as markers for head and neck malignancies. *Neoplasma* 1988, 35, 425-434.
- Gail MH, Muenz L, McIntire KR, et al. Multiple markers for lung cancer diagnosis: validation of models for advanced lung cancer. J Natl Cancer Inst 1986, 76, 805-816.
- 49. Straka MB, Wagner RL, Johnson JT, Kachman KK, Eibling DE. The lack of utility of a tumor marker panel in head and neck carcinoma: squamous cell carcinoma antigen, carcinoembryonic antigen, lipid-associated sialic acid, and CA-125. Arch Otolaryngol Head Neck Surg 1992, 118, 802-805.
- Dreyfuss AI, Clark JR, Andersen JW. Lipid-associated sialic acid, squamous cell carcinoma antigen, carcinoembryonic antigen, and lactic dehydrogenase levels as tumor markers in squamous cell carcinoma of the head and neck. Cancer 1992, 70, 2499-2503.
- Ropka ME, Goodwin J, Levine PA, Sasaki CT, Kirschner JC, Cantrell RW. Effective head and neck tumor markers: the continuing quest. Arch Otolaryngol Head Neck Surg 1991, 117, 1011-1014.
- 52. Mevio E, Benazzo M, Galioto P, Spriano P, Pizzala R. Use of serum markers in the diagnosis and management of laryngeal cancer. *Clin Otolaryngol* 1991, 16, 90-92.
- 53. Palermo F, Carniato A, Fede A, Boccaletto F, Marchiorie C.

- Serum SCC-AG in head and neck squamous cell carcinoma. Int J Biol Markers 1990, 5, 118-120.
- Screm MC, Grandis S, Cartei G, Cattaruzzi E. Detection of five circulating antigens in patients with head and neck squamous cell carcinoma. *Int J Biol Markers* 1989, 4, 35–39.
- Bhatavdekar JM, Gatel DD, Vora HH, Balar DB. Squamous cell carcinoma antigen and protein-bound sialic acid in the management of head and neck cancer. *Int J Biol Markers* 1991, 6, 237-240.
- Krecicki T, Leluk M. Acute phase reactant proteins-an aid to monitoring surgical treatment of laryngeal carcinoma. J Laryngol Otol 1992, 106, 613-615.
- Portoukalian J, David MJ, Shen X, Richard M, Dubreuil C. Tumor size-dependent elevations of serum gangliosides in patients with head and neck carcinomas. *Biochem Int* 1989, 18, 759-765.
- 58. Altissimi G, von Garrel C. Role of tumor marker TAG-72 in head and neck neoplasms. *HNO* 1990, **38**, 364–366.
- Rayo JI, Garcia-Talavera JR, Martin M, Munoz A, Del Canico A. Serum TATI levels and clinical correlation in tumors of the head and neck. Scand J Clin Lab Invest Suppl 1991, 207, 33-35.
- Negri L, Pacchioni D, Calabrese F, Giacomasso S, Mastromatteo V, Fazio M. Serum and salivary CEA and GICA levels in oral cavity tumors. Int J Biol Markers 1988, 3, 107-112.
- 61. Zoller J, Fiehn W, Mende U, Hotz G. The diagnostic value of the tumor markers CEA, "CA 19-9", "CA-125", "CA 15-3" and "SCC" for the detection of recurrent tumors in patients with tumors of the head and neck. Deutsch Zeitschrift Fur Mundkiefer und Gesichts-Chirurgie 1990, 14, 254-259.
- Lian SL, Hsu HY, Lin SM. Serum copper and zinc levels in patients with nasopharyngeal carcinoma. *Taiwan I Hsueh Hui Tsa Chih—J Formosan Med Assoc* 1989, 88, 236–239.
- Zeillinger R, Swoboda H, Machacek E, et al. Expression of cathepsin D in head and neck cancer. Eur J Cancer 1992, 28A, 1413-1415.

Acknowledgement—Supported in part by the Mary Hillman Jennings Foundation.